

Enzymatic branched ubiquitin chain formation on K63 homotypic chain with branched points defined by photo-induced stepwise poly-ubiquitin synthesis

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Protein ubiquitination is an essential post-translation modification that regulates numerous cellular processes. Ubiquitin modification in any of its seven lysine residues or N-terminal methionine of proximal ubiquitin results in polymeric chains. The structure complexity ranging from monoubiquitin, homotypic chain, and heterotypic chains with more than one linkage-type, gives rise to ubiquitin code and signal diversity¹. Even though recent studies have stated the importance of heterotypic chains, reports on its biological role or the mechanism involved in the synthesis are limited^{2,3}. Thus, the method enabling efficient synthesis of structure-defined ubiquitin chains is highly demanded for systematic studies on their functions.

To achieve the synthesis of structure defined polyubiquitin chains, a semi-synthetic method of step wise ubiquitin chain assembly was established. The proximally or distally modified/blocked monoubiquitins are used for the synthesis of structure-defined polyubiquitin chains by linkage defined ubiquitin ligase. The Ubiquitin with Nvoc-protected lysine at the 63rd position (K63) was synthesized by Solid phase peptide synthesis followed by native chemical ligation. Photo-induced deprotection of Nvoc-protected lysine residue on successive round of enzymatic reaction enables the synthesis of length-controlled K63 ubiquitin chains. Furthermore, blocking the acceptor site K48 by arginine substitution in ubiquitin moiety would limit the ubiquitination site with respect to branch point.

1) A.J. Boughton, K. Susan, F. David, *Structure*. **2019**, *28*, 29-43. 2) F. Ohtake, Y. Saeki, S. Ishido, J. Kanno, K. Tanaka, *Mol. Cell*, **2016**, *64*, 251-266. 3) L. Pluska, E. Jarosch, H. Zauber, A. Kniss, A. Waltho, K. Bagola, M. von Delbrück, F. Löhr, B.A. Schulman, M. Selbach, V. Dötsch, T. Sommer, *EMBO J.* **2021**, *40*, e106094.